

Bioaerosol sampling and collection techniques

Alexandra Tamm and Nina Ruckteschler
Max Planck Institute for Chemistry, Mainz/Germany
Bioaerosol class retreat in Bad Münster am Stein, Ebernburg, 12/13 Sep. 2016

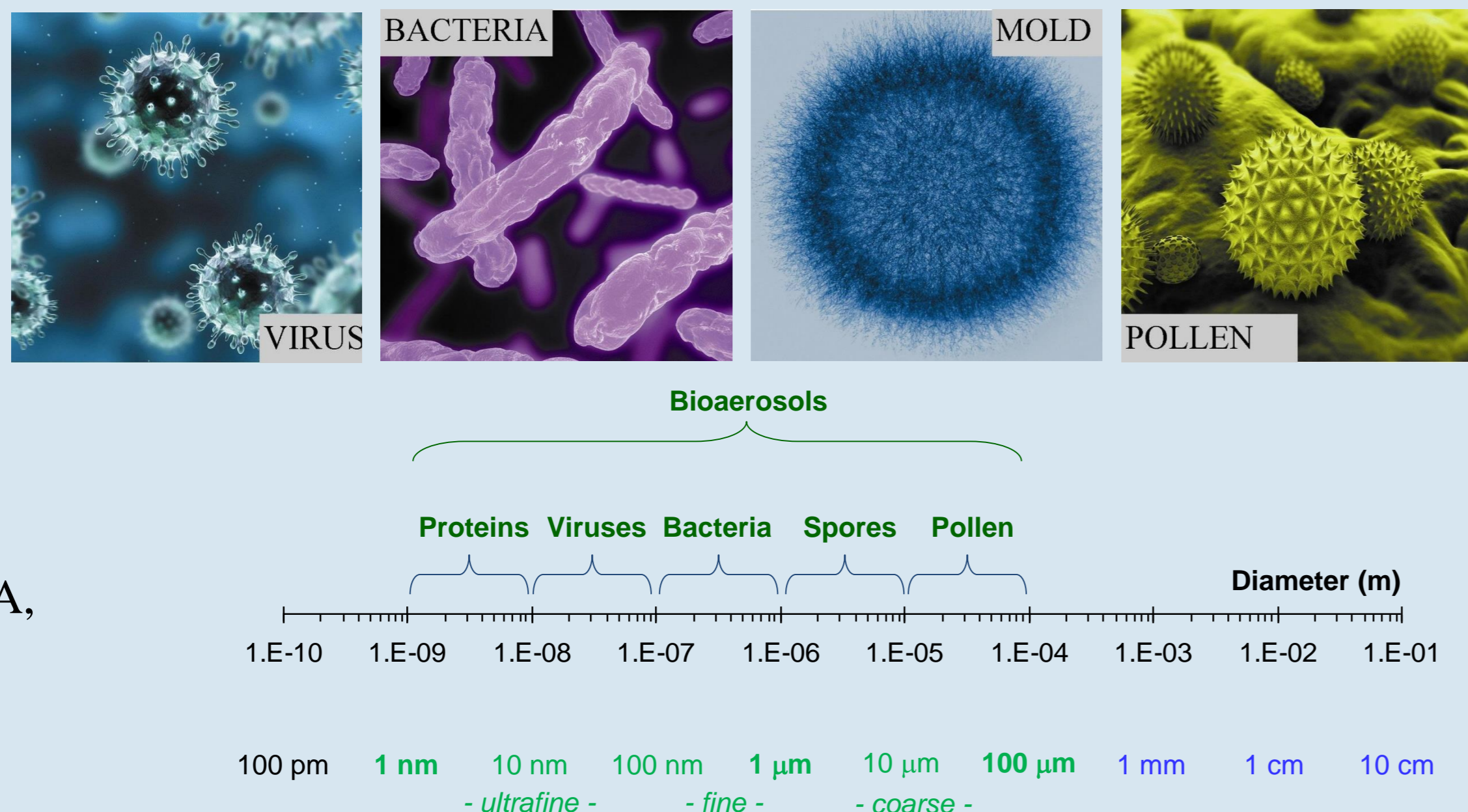


1. What are bioaerosols?

Bioaerosols (biological aerosols) are suspensions of airborne particles that contain or were released from living organisms.

Bioaerosols comprise pathogenic and non-pathogenic, living (mostly infectious?) and dead (mostly climate relevant?) material such as:

- viruses
- bacteria
- fungi, oomycetes
- algae
- cyanobacteria
- archaea
- lichens
- bryophytes
- vascular plants
- pollen and spores
- cell fractions, DNA, proteins, debris, excrements



2. Relevance of bioaerosol sampling

We inhale 10 m³ of air per day and in each m³ air are approx. **1000 cultivable microorganisms.**

Bioaerosols have influence on:

- health (especially respiratory tract and skin)
- agriculture and forestry
- precipitation: cloud condensation nuclei or ice nuclei → interaction with radiation → effect on climate

Sampling is influenced by:

- particle aerodynamic diameter
- wind velocity
- direction
- inlet characteristic
- sampling time
- mechanical stress
- desiccation

Keep in mind:

- various sampling techniques differ in detectable particle size range
- often pre-separator/pre filter are necessary
- What is the available budget for the sampling?

3. Methods

| Pro | Technique | Contra |
|---|---|--|
| <ul style="list-style-type: none"> • economically feasible • direct collection of microorganisms | <p>Impaction</p> <ul style="list-style-type: none"> • air is forced to change direction → larger particles hit the surface, smaller particles follow the airstream • collection on different kinds of substrate (e. g. glass slide, agar plate, filter, gelatine) • impactor types: virtual, slit and cascade | <ul style="list-style-type: none"> • quantification: culture based • overloading of culture plate → overlap of colonies • bounce off particles → transport in the next impaction stage • effect of wind speed • desiccation → low recovery efficiency |
| <ul style="list-style-type: none"> • liquid collection medium: reduction of overloading and desiccation stress of microorganism | <p>Impingement</p> <ul style="list-style-type: none"> • airstream is led into a liquid collection medium → microorganisms are collected by immersion | <ul style="list-style-type: none"> • quantification: post collection processes • evaporation of liquid medium → miscalculation of quantification • not compatible with size fractionation • no size fractionation |
| <ul style="list-style-type: none"> • economically feasible • potential for size fractionation • portable | <p>Filtration</p> <ul style="list-style-type: none"> • airstream is forced to go through different filters types of different materials (e. g. cellulose, glass, quartz, plastic) • filter types: fiber (A), membrane (B), porous foam (C), capillary pore (D) | <ul style="list-style-type: none"> • quantification: post collection processes • overloading of culture plate → overlap of colonies • desiccation → low recovery efficiency • effect of wind speed |
| <ul style="list-style-type: none"> • good collection efficiency → reduced particle bounce • low pressure drop (higher flow rates) | <p>Cyclone</p> <ul style="list-style-type: none"> • swirling air and centrifugal force is used to capture microorganisms into a liquid | <ul style="list-style-type: none"> • evaporation of liquid medium → miscalculation of quantification |
| <ul style="list-style-type: none"> • good recovery efficiency → reduced stress on microorganisms • quantification possible • no vacuum pump needed | <p>Electrostatic precipitation</p> <ul style="list-style-type: none"> • particles are charged at the inlet and exposed to an electrical field inside the sampler → particle migration over charged plate | <ul style="list-style-type: none"> • viability of bacteria is effected by electric charge • few study on this technique → less data for comparison |
| <ul style="list-style-type: none"> • good collection efficiency for smaller particles • determination of particle size distribution • air flows freely through the sampler → small pressure drop | <p>Thermal precipitation</p> <ul style="list-style-type: none"> • hot and cold surface → along a temperature gradient particles move to the cooler surface and will be collected (thermophoretic motion) | <ul style="list-style-type: none"> • low collection rate • small collection area • high temperature → effect on viability of the microorganisms |
| <ul style="list-style-type: none"> • ultrafine bioaerosol particles can also be sampled and detected easily • very low detection limit | <p>Condensation technique</p> <ul style="list-style-type: none"> • airstream is forced to go through a saturator → evaporated liquid condenses on particles → spectroscopical detection | <ul style="list-style-type: none"> • complex system → expertise required • high temperature → effect on viability of the microorganisms |
| <ul style="list-style-type: none"> • economically feasible • no vacuum pump needed | <p>Gravity</p> <ul style="list-style-type: none"> • agar plate/SEM plate → microorganisms from air settle on a plate | <ul style="list-style-type: none"> • not accepted by official guidelines • relies on air currents • weakly correlated with counts of quantitative methods (primary larger particle settle down, smaller stay in air longer) |

4. Artefacts

Definition: Physical, physicochemical and chemical changes / sources of errors, which cause a change of composition.

Positive artefacts: Filter material with a high surface activity can adsorb gas phase components (especially quartz fiber filters!).

Negative artefacts: Evaporation of aerosol particles due to pressure and temperature changes.

Chemical artefacts: Changes in composition through reactions, e.g. with oxidants.

5. Conclusion

Offline sampling: collection of aerosols
→ opportunity for cultivation and identification and often also for quantification

Online sampling: direct detection of number and size of particles.

THE sampling method suitable for collection of different types of bioaerosol **does not exist!**

It depends on the individual research question which sampling methods will be the best.

In most cases: **Combination of different (bio)aerosol detectors/collectors.**